



Pergamon

Tetrahedron Letters 41 (2000) 2599–2603

TETRAHEDRON
LETTERS

Combinatorial synthesis of trisaccharides via solution-phase one-pot glycosylation

Takashi Takahashi,* Masaatsu Adachi, Akihisa Matsuda and Takayuki Doi

*Department of Applied Chemistry, Graduate School of Science and Engineering, Tokyo Institute of Technology,
2-12-1 Ookayama, Meguro, Tokyo 152-8552, Japan*

Received 24 December 1999; revised 27 January 2000; accepted 28 January 2000

Abstract

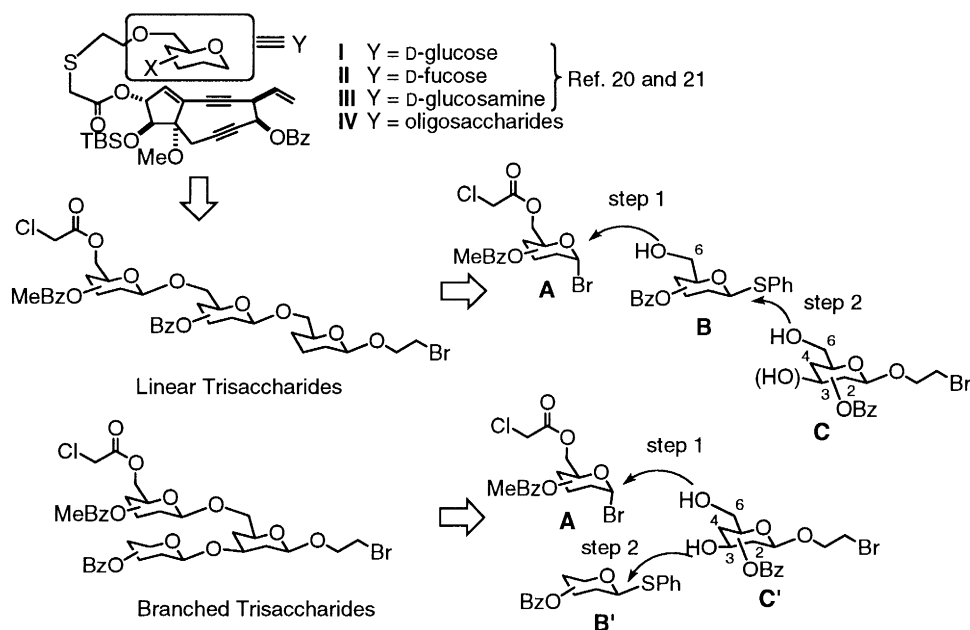
A library of 72 trisaccharides constructed from a combination of glucosides, galactosides, and mannosides via solution-phase one-pot glycosylation was synthesized rapidly using a manual synthesizer. © 2000 Elsevier Science Ltd. All rights reserved.

Keywords: glycosylation.

The combinatorial synthesis of oligosaccharides is a recent focus of our efforts in order to elucidate structure–activity relationships and to explore new bioactive molecules.^{1,2} Recently, several approaches for the synthesis of oligosaccharide libraries not only via solution-phase but also via solid-phase have been reported.^{3–13} We also reported that a one-pot, two step glycosylation is a potential method to rapidly assemble oligosaccharides.^{14–16} This simple method should prove beneficial for the synthesis of a library of oligosaccharides because the ordinal steps required for the elongation of glycosides, i.e., deprotection and purification of each synthetic intermediate, are omitted.^{17–19} Since we found that our reported DNA-cleaving enediyne molecules **I–III** containing mono-saccharides cleave DNA with certain sequence selectivity, such as 5′-CGG, 5′-CAG, and 5′-CGC, a variety of oligosaccharide-enediyne analogues **IV** should facilitate further studies in this area.^{20,21} Herein we wish to report the synthesis of a trisaccharide-library consisting of a combination of glucosides, galactosides, and mannosides by solution-phase one-pot glycosylation.

In our previous reports, a one-pot, two-step glycosylation was accomplished using a combination of different leaving groups, i.e., bromide (-Br) and phenylthio group (-SPh), with selective activators, AgOTf and NIS-TfOH, respectively, leading to linear- and branched-trisaccharides (Scheme 1).^{15,16} A library synthesis of linear-trisaccharides was examined through the combination of bromo glycosides **1–3**, phenylthio glycosides **4a–6a**, and 2-bromoethyl glycosides **7a–9a** or **7b–9b** (Fig. 1). The 2-bromoethyl group in **7–9** can be linked with the masked enediyne molecule to provide **IV** as previously

* Corresponding author.



Scheme 1. Strategy for a library synthesis of trisaccharides by a one-pot glycosylation

reported.²⁰ A chloroacetyl group can be utilized for the polymer-supported synthesis of a library of the masked enediyne molecules containing a variety of oligosaccharides for the next stage. In the synthesis of the linear trisaccharides, nine reactions were carried out simultaneously: bromo glucoside **1** was placed in nine Teflon tubes on a Quest 210[™] manual synthesizer.[†] Phenylthio glucoside **4a** was added to the first three vessels, phenylthio galactoside **5a** was added to the next three vessels, and

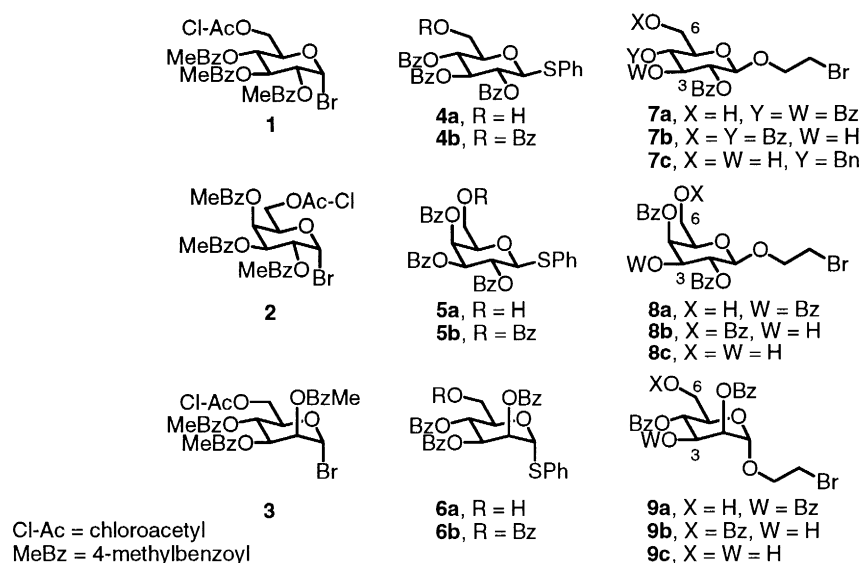


Fig. 1.

[†] A Quest 210 was purchased from Argonaut Technologies, USA.

phenylthio mannoside **6a** was added to the last three vessels. The mixtures were independently treated (AgOTf/CH₂Cl₂/0°C) in the presence of molecular sieves 4 Å. After all the reactions were complete as judged by TLC, 2-bromoethyl glycosides **7a–9a** or **7b–9b** were added to all three reaction vessels, respectively, as the combination of three components should be unique in all reaction vessels. Treatment of the reaction mixtures (NIS/cat. TfOH/0°C) provided the respective trisaccharides **10a–18a** or **10b–18b** in 64–99% yields after standard work-up and column chromatography. The results for all of the possible combinations for 54 trisaccharides are shown in Table 1. As anchoring is effected by the 2-position of the glycosyl donors, the anomeric linkages formed in the glycosylation were β-oriented (>95%) determined by the chemical shifts of ¹³C NMR (98.4–102.3 ppm), except that α-formation was observed when the mannosides were used as glycosyl donors (95.9–98.0 ppm).²²

Table 1
A library of linear 54 trisaccharides synthesized by one-pot glycosylation^{a,b}

-Br	-SPh	-O(CH ₂) ₂ Br	Product (Yield) ^c	-Br	-SPh	-O(CH ₂) ₂ Br	Product (Yield) ^c	
1	4a	7a or 7b	10a (83) 10b (74)	2	6a	7a or 7b	25a (77) 25b (79)	
		8a or 8b	11a (64) 11b (99)			8a or 8b	26a (78) 26b (86)	
		9a or 9b	12a (89) 12b (83)			9a or 9b	27a (90) 27b (63)	
	5a	7a or 7b	13a (91) 13b (79)		3	4a	7a or 7b	28a (79) 28b (77)
		8a or 8b	14a (73) 14b (73)				8a or 8b	29a (74) 29b (78)
		9a or 9b	15a (79) 15b (91)				9a or 9b	30a (91) 30b (97)
	6a	7a or 7b	16a (77) 16b (97)			5a	7a or 7b	31a (99) 31b (99)
		8a or 8b	17a (73) 17b (90)				8a or 8b	32a (86) 32b (93)
		9a or 9b	18a (82) 18b (99)				9a or 9b	33a (92) 33b (99)
2	4a	7a or 7b	19a (85) 19b (99)	6a	7a or 7b	34a (86) 34b (99)		
		8a or 8b	20a (64) 20b (95)		8a or 8b	35a (75) 35b (73)		
		9a or 9b	21a (78) 21b (82)		9a or 9b	36a (82) 36b (77)		
	5a	7a or 7b	22a (98) 22b (97)					
		8a or 8b	23a (75) 23b (99)					
		9a or 9b	24a (87) 24b (84)					

^a **1–3** (1.10 equiv.) and **4a–6a** (1.05 equiv.) were used for **7a–9a** (1.00 equiv.). ^b **1–3** (1.05 equiv.) **4a–6a** (1.00 equiv.) were used for **7b–9b** (1.00 equiv.). ^c Yields were calculated based on 2-bromoethyl glycosides **7–9**.

The one-pot glycosylation was also utilized in the synthesis of branched trisaccharides as follows: Glycosyl bromide **1–3** and phenylthio glycosides **4b–6b** were employed as glycosyl donors and 3,6-diol **7c–9c** were used as glycosyl acceptors (Fig. 1). The mixtures of a different combination of **1–3**, **4b–6b**, and 3,6-diol **7c**, placed in nine Teflon tubes on a Quest 210, were treated with AgOTf (step 1 in Scheme 1), followed by NIS/TfOH (step 2 in Scheme 1). By repeating this process, the desired 18 branched trisaccharides **37–54** were isolated in good yields (72–99%) after standard work-up and column chromatography (Table 2). When the galactoside **8c** was used as a glycosyl acceptor, the one-pot reactions failed because both of the hydroxy groups of **8c** had reacted with glycosyl bromides **1–3** in the first step.

We have demonstrated that a synthesis of a library of 72 trisaccharides with a combination of glycosides, galactosides, and mannosides can be accomplished by a one-pot, two step glycosylation, which proved to be a powerful method for the rapid synthesis of a library of trisaccharides using a manual

Table 2
A library of branched 18 trisaccharides synthesized by one-pot glycosylation^a

-Br	-SPh	-O(CH ₂) _n Br	Product (Yield) ^b	-Br	-SPh	-O(CH ₂) _n Br	Product (Yield) ^b
1	4b	7c	37 (93)	1	4b	9c	46 (95)
	5b	7c	38 (93)		5b	9c	47 (73)
	6b	7c	39 (78)		6b	9c	48 (72)
2	4b	7c	40 (97)	2	4b	9c	49 (99)
	5b	7c	41 (84)		5b	9c	50 (74)
	6b	7c	42 (99)		6b	9c	51 (87)
3	4b	7c	43 (99)	3	4b	9c	52 (99)
	5b	7c	44 (89)		5b	9c	53 (89)
	6b	7c	45 (99)		6b	9c	54 (99)
1-3	4b-6b	8c	- (0) ^c				

^a **1-3** (1.05 equiv.) and **4b-6b** (1.50 equiv.) were used for **7c-9c** (1.00 equiv.). ^b Yields were calculated based on 2-bromoethyl glycosides **7c-9c**. ^c Both hydroxy groups of 3,6-diol **8c** reacted with glycosyl bromides **1-3**.

synthesizer. Further study of library syntheses having other linkages, i.e., at the 2- and 4-positions of **C**, and the 2,6-, 4,6-, 2,3-, 2,4-, and 3,4-positions of **C'** instead of the 3,6-position (Scheme 1) is now underway in our laboratory.

Acknowledgements

This work was supported by a Grant-in-Aid from the Ministry of Education, Science, Sports and Culture, Japan (No. 11305065), Special Coordination Funds for Promoting Science and Technology from the Japanese Science and Technology Agency, and the Kurata Foundation (T.D.). The authors thank Professor H. Yamada for fruitful discussions.

References

- Schweizer, F.; Hindsgaul, O. *Curr. Opin. in Chem. Bio.* **1999**, *3*, 291–298.
- Arya, P.; Ben, R. N. *Angew. Chem., Int. Ed. Engl.* **1997**, *36*, 1280–1282 and references cited therein.
- Danishefsky, S. J.; McClure, K. F.; Randolph, J. T.; Ruggeri, R. B. *Science* **1993**, *260*, 1307–1309.
- Liang, R.; Yan, L.; Loebach, J.; Ge, M.; Uozumi, Y.; Sekanina, K.; Horan, N.; Gildersleeve, J.; Thompson, C.; Smith, A.; Biswas, K.; Still, W. C.; Kahne D. *Science* **1996**, *274*, 1520–1522.
- Kanie, O.; Barresi, F.; Ding, Y.; Labbe, J.; Otter, A.; Forsberg, L. S.; Ernst, B.; Hindsgaul, O. *Angew. Chem., Int. Ed. Engl.* **1995**, *34*, 2720–2722.
- Boons, G.-J.; Heskamp, B.; Hout, F. *Angew. Chem., Int. Ed. Engl.* **1996**, *35*, 2845–2847.
- Rodebaugh, R.; Joshi, S.; Fraser-Reid, B.; Geysen, H. M. *J. Org. Chem.* **1997**, *62*, 5660–5661.
- Öhrlein, R.; Baisch, G.; Katopodis, A.; Streiff, M.; Kolbinger, F. *J. Mol. Cat. B: Enzymatic* **1998**, *5*, 125–127.
- Rademann, J.; Geyer, A.; Schmidt, R. R. *Angew. Chem., Int. Ed. Engl.* **1998**, *37*, 1241–1245.
- Izumi, M.; Ichikawa, Y. *Tetrahedron Lett.* **1998**, *39*, 2079–2082.
- Nicolaou, K. C.; Watanabe, N.; Li, J.; Pastor, J.; Winssinger, N. *Angew. Chem., Int. Ed. Engl.* **1998**, *37*, 1559–1561.
- Zhu, T.; Boons, G.-J. *Angew. Chem., Int. Ed. Engl.* **1998**, *37*, 1898–1900.
- Wong, C.-H.; Ye, X.-S.; Zhang, Z. *J. Am. Chem. Soc.* **1998**, *120*, 7137–7138.

14. Yamada, H.; Harada, T.; Takahashi, T. *J. Am. Chem. Soc.* **1994**, *116*, 7919–7920.
15. Yamada, H.; Harada, T.; Miyazaki, H.; Takahashi, T. *Tetrahedron Lett.* **1994**, *35*, 3979–3982.
16. Yamada, H.; Kato, T.; Takahashi, T. *Tetrahedron Lett.* **1999**, *40*, 4581–4584.
17. Raghavan, S.; Kahne, D. *J. Am. Chem. Soc.* **1993**, *115*, 1580–1581.
18. Zhang, Z.; Ollmann, I. R.; Ye, X.-S.; Wischnat, R.; Baasov, T.; Wong, C.-H. *J. Am. Chem. Soc.* **1999**, *121*, 734–753 and references cited therein.
19. Tsukida, T.; Yoshida, M.; Kurokawa, K.; Nakai, Y.; Achiha, T.; Kiyoi, T.; Kondo, H. *J. Org. Chem.* **1997**, *62*, 6876–6881.
20. Takahashi, T.; Tanaka, H.; Matsuda, A.; Doi, T.; Yamada, H. *Bioorg. Med. Chem. Lett.* **1998**, *8*, 3299–3302.
21. Takahashi, T.; Tanaka, H.; Matsuda, A.; Doi, T.; Yamada, H.; Matsumoto, T.; Sasaki, D.; Sugiura, Y. *Bioorg. Med. Chem. Lett.* **1998**, *8*, 3303–3306.
22. Kotowycz, G.; Lemieux, R. U. *Chem. Rev.* **1973**, *73*, 669–698.